



THE DIAGNOSTIC PARASITOLOGY LABORATORY

Tel: +44 (0)20 7927 2427 www.parasite-referencelab.co.uk

ACANTHAMOEBA SPECIMEN REFERRAL FORM

Patient Surname	Other names	
Date of Birth	m/f	
NHS no. (or Hosp no.)	Lab no.	
Name and address of doctor referring specimen:	ESSENTIAL: telephone number for contact relating to this specimen	
Name and address for reports (if different):	Name and address for invoices (if different):	
	Purchase order number:	
Type of specimen: (please tick as appropriate)	Investigation required:	
RightLeftCorneal scrape / biopsy	Culture + PCR	
Corneal swab	Culture only	
Contact lens		
Other (please specify below):	Specimen date:	
Clinical details:		
Please give details of antimicrobial treatment: Please return form and specimen by post/courier to The Diagnostic Parasitology Laboratory. Faculty of Infectious & Tropical		

Please return form and specimen by post/courier to The Diagnostic Parasitology Laboratory, Faculty of Infectious & Tropical Diseases, London School of Hygiene & Tropical Medicine, Keppel Street, London WC1E 7HT OR via Hays DX to HPA Malaria Reference Lab, DX 6641200, Tottenham Crt RD92WC. SEE OVERLEAF FOR SPECIMEN REQUIREMENTS / GUIDELINES

Guidelines for referral of specimens to the Diagnostic Parasitology Laboratory at the London School of Hygiene and Tropical Medicine for the laboratory diagnosis of *Acanthamoeba* keratitis (AK)

All specimens should be submitted for testing together with a completed **Acanthamoeba DPL referral form** See previous page, and available to download and print from: <u>www.parasite-referencelab.co.uk</u>

The highest laboratory diagnostic sensitivity for AK is achieved through use of <u>both</u> *in vitro* culture / microscopy and a probe-based qPCR.

<u>Contact</u>: please telephone the Laboratory with queries, or for advice: 020 7927 2427.

Suitable sample types & sample preparation:

• Clinical samples: the ideal and preferred sample for diagnosis of AK is a corneal scrape / biopsy. The material should be collected with a needle or blade and rinsed into a small volume (1 - 2mL) of sterile saline / sterile distilled water in a small (<5mL) sterile vial / tube. Remove blades or needles as soon as possible after rinsing and before sending. Please do NOT leave blades in the tube as they rust: rust inhibits our PCR and may also have a detrimental effect on culture isolation.

Other clinical sample types:

- Corneal swabs. Do not send dry swabs: please add a small volume (1 2mL) of sterile saline or sterile distilled water to the swab to prevent drying.
- Corneal fluids, washings etc. should be sent in a small (<5mL) sterile vial / tube
- Punch biopsies or portions of excised cornea may also be submitted: put sample into a small volume of sterile saline / distilled water in a small sterile vial.
- Non-clinical samples. Contact lenses should be sent in their lens cases i.e. in used contact lens fluid. Culture is performed on lenses and associated fluid; PCR is performed on fluid only. NB: isolation of *Acanthamoeba* from contact lens-related specimens, whilst suggestive, does not prove the amoeba as causing the patient's symptoms. <u>Please note</u>: we do not test commercial contact lens solutions (other than that already in the patients' contact lens case).
- **Culture-positive sample isolates for confirmation / identification**: please send original culture plate, if possible, or blocks of agar from the plate in a sterile vial.

Please ensure all containers are tightly screwed and use Parafilm (NOT Sellotape) to prevent leakage during transit.

Specimen turnaround time (TAT): is 2 - 10 working days. Positive results from either culture or qPCR method will be telephoned in advance of the final result.

Sending samples:	by post / courier:	by Hays DX:
	Diagnostic Darasitology Laboratory	HDA Malaria Reference La

LSHTM Keppel Street, LONDON WC1E 7HT HPA Malaria Reference Lab TOTTENHAM CRT RD92WC DX 6641200